

**JIWAJI UNIVERSITY, GWALIOR-474011**  
**M.SC. BIOCHEMISTRY**

**CURRICULUM - 2010-2012**

Semester	Title of the Paper	Marks	
		EA	IA
<b>First</b>	101. Cell Biology	35	15
	102. Biomolecules	35	15
	103. Microbiology	35	15
	104. Tools and Techniques	35	15
	105. Lab Course I	35	15
	106. Lab. Course II	35	15
<b>Second</b>	201. Molecular Biology	35	15
	202. Immuno Technology	35	15
	203. Enzyme Technology	35	15
	204. Bioenergetics and Metabolism	35	15
	205. Lab Course III	35	15
	206. Lab Course IV	35	15
<b>Third</b>	301. Genetic Engineering	35	15
	302. Plant Biochemistry	35	15
	303. Animal Physiology & Endocrinology	35	15
	304. Nutritional and Clinical Biochemistry	35	15
	305. Lab Course V	35	15
	306. Lab Course VI	35	15
<b>Fourth</b>	401. Frontier Technologies in Biosciences	35	15
	402. Biostatistics, Computers and Bioinformatics	35	15
	403 Lab Course VII	35	15
	404 Project Work*	100	50

\*Evaluated both by the Internal & External examiner at the time of presentation.  
There are weekly seminars and continuous internal assessment throughout the course.

## 101: CELL BIOLOGY

### UNIT I

1. Cell Membrane: Physicochemical Properties; Molecular Organization – asymmetrical organization of lipids, proteins and carbohydrates; Biogenesis and Functions
2. Transport of Small Molecules Across Cell Membranes: Types and Mechanism
3. Active Transport by ATP-Powered Pumps: Types, properties and mechanisms
4. Transport of proteins into mitochondria and chloroplast

### UNIT II

1. Transport of proteins into and out of nucleus
2. Transport of proteins into endoplasmic reticulum
3. Transport by vesicle formation: Endocytosis and Exocytosis
4. Molecular Mechanism of vesicular transport

### UNIT III

1. Intracellular Digestion: Ultra structure and Functions of Lysosomes
2. Cell Motility and Shape I: Structure and Functions of Microfilaments
3. Cell Motility and Shape II: Structure and Functions of Microtubules and Intermediate Filaments
4. Intracellular communication through cell junctions: Occluding junctions, Anchoring Junctions and Communicating Junctions

### UNIT IV

1. Molecular Mechanism of Cell-Cell Adhesion:  $Ca^{++}$  dependent cell-cell adhesion
2. Molecular Mechanism of Cell-Cell Adhesion:  $Ca^{++}$  independent cell-cell adhesion
3. Extra-cellular Matrix of Animals: Organization and Functions
4. Extra-cellular Matrix Receptors on Animal Cells: The Integrins

### UNIT V

1. Cell Signaling: Signaling via G-Protein linked and enzyme linked cell surface receptors, MAP Kinase Pathways, Interaction and Regulation of signaling pathways
2. Eukaryotic Cell Division Cycle: Different Phases and Molecular Events
3. Control of Cell Division Cycle: In yeast and mammalian cells
4. Apoptosis: Phases and significance, Morphological and Biochemical changes associated with apoptotic cells, Apoptotic Pathways and regulators

### Practical Exercises

1. Sub cellular fractionation
2. Chromosome Preparation: Mitosis – Onion root tip, rat/mouse cornea, rat/mouse bone marrow, human lymphocytes
3. Chromosome Preparation: Meiosis – Rat/mouse testis, Grasshopper testis
4. Polytene chromosome preparation from *Drosophila* salivary gland
5. Identification of tissue typing: Histological preparation of tissue
6. Identification of different biomolecules in different tissues by histochemical techniques
7. Electron microscopy: Demonstration and good photographs for interpretation.

## **Reference Books**

1. Molecular Biology of the Cell (2002), Alberts, et al
2. Molecular Cell Biology (2004), Lodish, et al
3. Working with Molecular Cell Biology: A study Companion (2000), Storrie et al
4. Cell and Molecular Biology: Concepts and Experiments (3<sup>rd</sup> Ed., 2002), Gerald Karp
5. The Cell: A Molecular Approach (2004), G.M. Cooper
6. The Word of the Cell (1996), Becker et al
7. Cell Proliferation and Apoptosis (2003), Hughes and Mehnet
8. Essential Cell Biology (1998), Alberts et al
9. Biochemistry and Molecular Biology of Plants (2000), Buchanan et al
10. Harpers Biochemistry Murray et al

*Note:* All text books are of latest editions.

## 102 : BIOMOLECULES

### UNIT I

1. Carbohydrates : Structure, classification, properties and functions
2. Home and heteropolysaccharides : carbohydrate derivatives
3. Lipids : Classification, structure, properties and functions
4. Lipids with special biological functions

### UNIT II

1. Amino acids : Structure, classification, abbreviations, properties and functions
2. Peptides and polypeptides
3. Synthesis of peptides and protein sequencing
4. Proteins : Properties, covalent structure, secondary, tertiary and quaternary structure

### UNIT III

1. Enzymes : Classification, mechanism of action, allosteric enzymes, multienzyme complex
2. Enzyme kinetics : Basic concepts
3. Water soluble vitamins : Structure, distribution, interaction and biological functions (mechanism of action not included)
4. Fat soluble vitamins : Structure, distribution and functions

### UNIT IIII

1. Nucleotides : Structure of purine and pyrimidine bases, nucleosides, nucleotides
2. DNA : Structure and Conformation
3. DNA : denaturation, degradation, modification, repair, recombination and rearrangement
4. RNA : Structure, types and functions

### UNIT V

1. Animal hormones : Structure and biological roles
2. Plant hormones : Structure and biological functions
3. Plant phenolics: Classification and functions
4. Alkaloids : Classification and functions

### Practical Exercises

1. Titration of amino acids
2. Colorimetric determination of pKa
3. Model building using space filling/ ball and stick models
4. Reaction of amino acids, sugars and lipids
5. Quantitation of proteins and sugars
6. Analysis of oils : iodine number, saponification value, acid number

### Reference Books

1. Principles of Biochemistry by Nelson, Cox and Lehninger
2. Biochemistry by G.Zubay
3. Biochemistry by Stryer
4. Biochemistry by Garrett and Grisham

5. Biochemical Calculations, Irwin H. Seigel, John Wiley and Sons Inc.
6. Biochemistry, DVoet and JG. Voet , J Wiley and Sons.
7. Biochemistry, D Freifilder, W.H. Freeman & Company.
8. Laboratory Techniques in Biochemistry and molecular Biology, Work and Work
9. A Biologist's guide to Principles and Techniques of Practical Biochemistry, Wilson & Goulding, ELBS Edition.

*Note:* All texts are of latest editions.

## 103 : MICROBIOLOGY

### UNIT I

1. Classification of Microorganisms: Basis of microbial classification, Haeckel three kingdom , Whittaker's five kingdom concept.
2. Morphology and fine structure of eubacteria and archeobacteria cell wall, cytoplasmic membrane and other organelles.
3. Pure culture techniques and preservation methods.
4. Preparation of Culture media, microbial staining.

### UNIT II

1. Sterilization: Physical and chemical methods
2. Microbial Growth: Bacterial growth curve, Mathematical expression, measurement of Growth and factors affecting growth
3. Microbial Nutrition: Nutritional classification of Microorganisms, common nutritional requirements, mode of nutrition, transport of nutrients across the bacterial membrane
4. Oxygen toxicity: Study of catalase, peroxidase, superoxide dismutase, mechanism of oxygen toxicity.

### UNIT III

1. Virus: Types, Isolation, cultivation, identification and viral replication.
2. Structure and morphology of Bacteriophage, Lytic and lysogenic cycle.
3. Life cycle of DNA Viruses: SV 40, RNA Viruses: Retroviruses
4. Cyanobacteria : General account and their importance

### UNIT IV

1. Infection and disease, types of Infection, Mechanism of pathogenicity
2. Bacterial Diseases: Staphylococcal and Clostridial food poisoning, Salmonellosis Shigellosis
3. Fungal diseases: Histoplasmosis , Aspergillosis
4. Viral diseases: Chicken pox, Hepatitis B, and Poliomyelitis

### UNIT V

1. Mycoplasmas and diseases caused by them
2. Bacterial Recombination : Transformation, Conjugation, Transduction, Plasmids and transposons
3. Chemotherapeutic agents : Classification of antibiotics, Broad spectrum antibiotics; Antibiotics from prokaryotes
1. Anti-fungal and antiviral antibiotics, mode of action of antibiotics and resistance to antibiotics

### Practical Exercises

1. Preparation of liquid and solid media for growth of microorganisms.
2. Isolation and maintenance of organisms by plating, streaking and serial dilution methods. Slants and stab cultures. Storage of microorganisms.
3. Isolation of pure cultures from soil and water.
4. Growth; Growth curve ; Measurement of bacteria population by turbidometry and serial dilution methods. Effect of temperature, pH and carbon and nitrogen

sources on growth.

5. Microscopic examination of bacteria, Yeast and molds and study of organisms by Gram stain, Acid fast stain and staining for spores.
6. Study of mutations by Ames test.
7. Assay of antibiotics and demonstration of antibiotic resistance.
8. Analysis of water for portability and determination of MPN.
9. Bacterial transformation.
10. Biochemical characterization of selected microbes.
11. One step growth curve of coliphage.

### **Reference Books**

1. General Microbiology, R.Y. Ingraham, J.L. Wheelis, M.L. and Painter, P.R. The MacMillan Press Ltd.
2. Brock Biology of Microorganism, M.T. Martinko, J.M. and Parker, J. Prentice-Hall.
3. Microbiology, Pelzar, M.J., Chan , E.C.S. and Kreig, N.R., Tata McGraw Hill.
4. Microbial Genetics, Maloy , S.R., Cronan, J.E.Jr and Freifelder, D. Jones, Bartlett Publishers.
5. Microbiology- a Laboratory Manual, cappuccino, J.G. and Sherman, N. Addison Weseley.
6. Microbiological Applications, (A Laboratory Manual in General Microbiology) Benson, H.J. WCB: Wm C. Brown Publishers

Note: All text books are of latest editions

## 104 : TOOLS AND TECHNIQUES

### UNIT I

1. Centrifugation : Basic principle, type, instrumentation and applications
2. Photometry : Basic principles of colorimetry, and UV visible spectrophotometry, instrumentation and applications
3. Infra red spectroscopy
4. Fluorimetry : Principle, instrumentation and applications

### UNIT II

1. Chromatography : Principle, types, instrumentation and applications
2. Affinity chromatography, HPLC and FPLC
3. Electrophoresis : Principle, types and applications
4. Isoelectric focussing and isotachopheresis

### UNIT III

1. Atomic absorption spectroscopy : Principle, instrumentation and applications
2. Flame emission spectroscopy : Principle, instrumentation and applications
3. Polarimetry : Principle, instrumentation and applications
4. ORD and CD

### UNIT IV

1. ESR : Principle, instrumentation and applications
2. NMR, GC Mass : Basic principle, instrumentation and applications
3. X ray crystallography : Principle, instrumentation and applications
4. Radio immunoassay : Basic principle and applications

### UNIT V

1. Microscopy : Light, phase contrast, interference, fluorescence and polarization microscopy
2. Electron microscopy : Principle and Applications
3. Radioactivity : Principle, Geiger Muller Counter, liquid scintillation counter, solid scintillation counter, gamma counter
4. Autoradiography : Principles, and applications

### Practical Exercises

1. Verification of Beer's law
2. Determination of absorption maxima
3. Electrophoresis of Proteins- native and under denaturing conditions.
4. Amino acid and carbohydrate separations by paper & thin layer chromatography
5. Gas chromatography
6. Ion exchange and gel filtration chromatography
7. Separation of blood cells by density gradient centrifugation

### Reference Books

1. Physical Biochemistry : Applications to Biochemistry and Molecular Biology by Freifelder
2. Biochemical Techniques : Theory and Practice by Robyt and White

3. Principles of Instrumental Analysis by Skoog and West
4. Analytical Biochemistry by Holme and Peck
5. Biological Spectroscopy by Campbell and Dwek
6. Organic Spectroscopy by Kemp
7. A Biologist's Guide to Principles and Techniques of Practical Biochemistry by Wilson and Goulding
8. Principles of Instrumental Analysis by Skoog, Hollar and Nicman

*Note:* All text books are of latest editions.

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### **105 : Lab Course I**

Consists of practical exercises listed out under 101 & 102

### **106 : Lab Course II**

Consists of practical exercises listed out under 103 & 104

## 201: MOLECULAR BIOLOGY

### UNIT I

1. Nature of Gene : Evolution of Gene Concept, Chemical Nature of Gene, Gene-cistron relationship in prokaryotes and eukaryotes
2. DNA Replication : General features of Chromosomal Replication; DNA Replication Machinery in Prokaryotes and its comparison with Eukaryotes
3. Enzymology of DNA Replication : DNA Polymerases; Primases; Ligases; Helicases; Topoisomerases; Gyrase and Single Stranded Binding Proteins
4. Regulation of DNA Replication

### UNIT II

1. Transcription in Prokaryotes : Initiation, elongation and termination
2. Structure and functions of prokaryotic promoter
3. Control of transcriptional initiation in prokaryotes : Structure and functions of RNA Polymerase; Sigma factors – Types and functions
4. Control of transcriptional termination in prokaryotes : Intrinsic termination and Rho factor dependent termination; attenuation and antitermination

### UNIT III

1. Regulation of Gene Expression in Prokaryotes : Operon concept, induction and repression, Structure and regulation of lactose, arabinose and tryptophan operons
2. Initiation of Transcription in Eukaryotes : RNA Polymerases – Types and properties; Promoter – Types, structure and properties
3. Transcription factors – Types and properties; Enhancers – Structure and properties; Response Elements
4. Post-transcriptional Modification Eukaryotes – 5' and 3' modification of mRNA

### UNIT IV

1. Post-transcriptional Processing of pre mRNA, pre rRNA and pre tRNA transcripts
2. Genetic Code : Evidence and properties; Wobble hypothesis; Transcriptional adaptors and amino acyl tRNA synthetases
3. Translation : Successive stages of protein synthesis in prokaryotes and its comparison with eukaryotes
4. Post-translational Modification : Types and Significance

### UNIT V

1. Regulation of Gene Expression in Eukaryotes: *cis*-acting DNA Elements; Chromatin organization and regulation of gene expression; Regulation at the level of processing of transcripts
2. Regulation of Gene Expression in Eukaryotes : RNA Editing; Gene Alteration; DNA methylation and gene regulation; Regulation of gene expression by hormones; Regulation of gene expression at translational level
3. Transposable elements in Prokaryotes and Eukaryotes: Types and significance
4. Oncogenes and Tumor Suppressor Genes: Properties and significance

### **Practical Exercises**

1. Isolation of genomic DNA and restriction digestion
2. Size fractionation of restricted DNA fragments by Agarose Gel Electrophoresis
3. Quantitation of DNA
4. Determination of Amax of purified DNA fragments
5. Determination of Tm of nucleic acid
6. Isolation of RNA
7. Fractionation of poly (A)<sup>+</sup> RNA
8. *In vitro* transcription
9. *In vitro* translation
10. Metabolic labeling of proteins and immunoprecipitation
11. Protein- DNA interaction

### **Reference Books**

1. Genes VIII , Benjamin Lewin
2. Molecular Biology , Turner et al
3. Cell and Molecular Biology : Concepts and Experiments , Gerald Karp
4. Transcriptional Regulation in Eukaryotes (2000), Carey and Smale
5. Translational control of gene Expression (2000), Sonenberg et al
6. Chromatin and Gene Regulation (2001), Turner
7. An Introduction to Genetic Analysis , Griffiths et al
8. Genome (1999), Brown
9. Concepts of Genetics, Klug and Cummings
10. Proteins , Creighton
11. Molecular Cell Biology , Lodhish et al
12. Biochemistry and Molecular Biology of Plants (2000), Buchanan
13. Plant Biochemistry and Molecular Biology , Lea and Leegood
14. Plant Biochemistry (1997), Dey and Harborne

*Note:* All text books are of latest editions.

## 202 : IMMUNOTECHNOLOGY

### UNIT I

1. Immune response : Innate immune mechanisms and characteristics of adaptive immune response, Hematopoiesis
2. Anatomical organization of immune system : Primary lymphoid organs, Secondary lymphoid organs. Ontogeny and Phylogeny of lymphocytes, Lymphocyte traffic
3. Cell of the immune system : Mononuclear cells and granulocytes, Antigen presenting cells, Lymphocytes and their subsets. Antigens, Heptanes: Factors affecting immunogenicity, properties of T and B Cell epitopes, Super antigens
4. Inflammation; its mediators and the process, Cell adhesion molecules and their role in inflammation, lymphocyte homing, tissue injury and immune response leading to an inflammatory reaction, role of anaphylatoxins, granulocytes in inflammatory process

### UNIT II

1. Major histocompatibility systems : Structure of MHC I and II molecules, polymorphism, distribution variation and function. Organization of MHC complex in Mouse and Humans. Association of MHC with disease
2. Recognition of antigens by T and B Cells : Antigen processing, Role of MHC molecules in antigen presentation and Costimulatory signals
3. T – cell receptor complex, T-cell accessory membrane molecules, activation of T cells, Organization and arrangement of T-receptor genes
4. B-cell receptor complex, Activation of B-cells, Immunoglobulins : Molecular structure, types and functions. Antigenic determinants on immunoglobulins

### UNIT III

1. Molecular mechanism of antibody diversity : Organization of genes coding for constant and variable regions of heavy chains and light chains. Mechanisms of antibody diversity, Class switching
2. Antibody engineering, Antigen-Antibody interaction, avidity & affinity measurement
3. Monoclonal antibodies: Production, characterization and applications in diagnosis, therapy and basic research
4. Complement system, components, activation pathways, and regulation of activation pathways, Complement deficiencies, Role of complement system in immune responses

### UNIT IV

1. Cytokines : Structure and functions, cytokine receptors, signal transduction mediated by cytokine receptors, cytokine regulation of immune responses, cytokine related diseases and therapeutic applications of cytokines
2. Cytotoxic T cells and their mechanism of action, NK cells and mechanism of target cell destruction. Antibody dependent cell mediated cytotoxicity, Delayed type hypersensitivity. Techniques of Cell mediated immunity
3. Immunoregulation mediated by antigens, antibodies, immune complexes, MHC and cytokines
4. Hypersensitivity : Definition, IgE mediated hypersensitivity, mechanism of mast cell degranulation, mediators of type- I reactions and consequences. Type II reactions, Immune complex mediated hypersensitivity and Delayed type hypersensitivity

## **UNIT V**

1. Autoimmunity : Organ specific diseases, Systemic diseases, Mechanisms of autoimmunity and therapeutic approaches
2. Immunodeficiency syndromes : Primary immunodeficiencies and Secondary immunodeficiencies and their diagnosis and therapeutic approaches
3. Vaccines : Active and passive immunization, Whole organism vaccines, Macromolecules as vaccines, Recombinant-vector vaccines, DNA Vaccines, Synthetic peptide vaccines and sub-unit vaccines
4. Immunodiagnostics : Precipitation techniques, Agglutination, Fluorescence techniques, ELISA, RIA, Western blotting and Immno-histochemical techniques

### **Practical Exercises**

1. Blood Film preparation and identification of cells.
2. Lymphoid organs and their microscopic organization.
3. Immunization and production of polyclonal antibodies
4. Double diffusion and Immuno- electrophoresis.
5. Radial Immunodiffusion.
6. Purification of IgG from serum.
7. Separation of mononuclear cells by Ficoll- Hypaque.
8. Con-A induced proliferation of thymocytes (by MTT method).
9. Western –blotting.
10. ELISA
11. Preparation of antibody-enzyme conjugates

### **Reference Books**

1. Kubey, Immunology, R.A. Goldsby, Thomas J. Kindt, Barbara, A.Osbarne.(Freeman).
2. Immunology- Ashort Course, -Eli Benjamini, Richard Coico, Geoffrey Sunshine.
3. Immunology by Tizzard
4. Fundamentals of immunology, William Paul.
5. Immunology, by Roitt and others.
6. Immunology by Abbas

*Note:* All text books are of latest editions.

## 203 : ENZYME TECHNOLOGY

### UNIT I

1. Enzyme : Historical aspects, classification and nomenclature, EC number.
2. Mechanism of enzyme catalysis and action
3. Sub cellular localization and organization of enzymes
4. Methods of enzyme assay: continuous and sampling techniques, coupled enzyme assay and methods and significance of enzyme turnover number; specific activity

### UNIT II

1. Enzyme purification techniques : objectives and strategy; methods of homogenization; method of isolation; purification and crystallization
2. Criteria of purity and tabulation of purification data; stable storage of enzymes
3. Characterization of purified enzyme
4. Coenzymes, Cofactors and Isoenzymes

### UNIT III

1. Enzyme Kinetics : Equilibrium and steady state theory, rate equation and determination of  $K_m$  and  $V_{max}$
2. Factors affecting rate of enzyme reaction: pH, temperature and pressure
3. Enzyme inhibition : reversible and irreversible inhibition, their type, inhibitor constant and its significance
4. Rapid reaction techniques

### UNIT IV

1. Protein- ligand binding : types, cooperativity, Hill and Scatchard plot, Allosteric enzymes : Models of allostery, types and kinetics
2. Regulation of enzymes
3. Mechanism of action of Chymotrypsin; Ribonuclease; Lysozyme; Metallo-enzymes
4. Degradation of enzymes

### UNIT V

1. Enzyme immobilization; techniques; experimental procedures and effect of immobilization on kinetic parameters
2. Principle and Industrial application of immobilized systems
3. Enzymes in Medical diagnosis and enzyme therapy
4. Enzymes during aging

### Practical Exercises :

1. Protein estimation methods: Lowry, Bradford and Spectrophotometric.
2. Urease estimation in plant tissues
3. Assay of Acid phosphatase in plant seeds
4. Assay of Alkaline phosphatase in Kidney and Liver
5. Determination of optimum pH, temperature & time
6. Determination of  $K_m$  value of alkaline phosphatase
7. Acetylcholinesterase estimation in Rat /Goat Brain
8. Enzyme purification: Ammonium sulphate precipitation, Ion exchange chromatography , molecular sieve chromatography.

9. Checking of purity of enzyme by PAGE
10. Molecular weight determination of enzyme by Gel Filtration
11. Immobilization of HRP (Horse reddish peroxidase).
12. Kinetic properties of Immobilised HRP
13. Sub-cellular fractionation of rat liver and marker enzyme assays.

### **Reference Books**

1. The Nature of Enzymology by R.L. Foster
2. Enzymes by Dixon and Webb
3. Fundamentals of Enzymology by Price and Stevens
4. Enzyme Catalysis and Regulation by Hammes
5. Enzyme Reaction Mechanisms by Walsch
6. The Enzymes vol I and II by Boyer
7. Enzyme Structure and Mechanism by Alan Fersht
8. Enzyme Assays: A Practical Approach by Eisenthal and Danson
9. Enzyme Biotechnology by G. Tripathi
10. Practical Biochemistry by Plummer.
11. Practical Biochemistry by Sawhney and R. Singh

*Note:* All text books are of latest editions.

## 204 : BIOENERGETICS AND METABOLISM

### UNIT I

1. First and second laws of thermodynamics
2. Concept of free energy
3. ATP Cycle, ATP as high energy compound, functions of ATP
4. Other high energy biological compounds

### UNIT II

1. Basic Concepts of intermediary metabolism
2. Carbohydrate metabolism : Glycolysis, Kreb's cycle, glycogenolysis, glycogenesis, pentose phosphate pathway, gluconeogenesis, glyoxalate pathway
3. Regulation of carbohydrate metabolism
4. Inborn errors of carbohydrate metabolism

### UNIT III

1. Electron transport and oxidative phosphorylation
2. Biosynthesis and degradation of lipids
3. Regulation of lipid metabolism
4. Inborn errors of lipid metabolism

### UNIT IV

1. Nitrogen assimilation
2. Biosynthesis of amino acids
3. Degradation of amino acids
4. Regulation of amino acid metabolism

### UNIT V

1. Inborn errors of amino acid metabolism
2. Nucleic acid metabolism
3. Inborn errors of nucleic acid metabolism
4. Integration of metabolism

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### 205 : Lab Course III

Consists of practical exercises listed out under 201 & 202

### 206 : Lab Course IV

Consists of practical exercises listed out under 203 & 204

## 301 : GENETIC ENGINEERING

### UNIT I

1. The recombinant DNA Technology : General concept and Principle of cloning
2. Enzymes : Nucleases and restriction endonucleases – properties and types; phosphomonoesterases; polynucleotide kinase; DNA ligase; DNA polymerase I; RNA Dependent DNA Polymerase; terminal deoxynucleotidyl transferase; poly A polymerase
3. Prokaryotic host-vector system : Characteristics of *E. coli* as host; vectors for cloning in *E. coli* (plasmid, bacteriophage and plasmid-phage)
4. Other Prokaryotic host vector systems : Characteristics of Gram positive and Gram negative organisms as host and suitable vectors for cloning; Shuttle Vectors

### UNIT II

1. Design and characteristics of expression vectors for cloning in prokaryotes
2. Factors affecting expression of cloned genes in prokaryotes
3. Cloning in Yeast : Properties of yeast as host for cloning and different types of vectors designed for cloning in yeast
4. Cloning in animals system : Animal system as a model host, Methods of introduction of foreign DNA in animal system; Vectors for cloning in animal system – SV 40, bovine papilloma virus, adenovirus, vaccinia virus, baculovirus and retrovirus vectors

### UNIT III

1. Methods for Constructing rDNA and cloning: Inserts; vector insert ligation; infection, transfection and cloning
2. Methods for screening and selection of recombinant clones
3. DNA Libraries: Types, advantages and disadvantages of different types of libraries; Different methods for constructing genomic and full length cDNA libraries
4. Gross anatomy of cloned insert – size, restriction mapping and location

### UNIT IV

1. Fine anatomy of DNA segment – General principle of chemical and enzymatic methods of nucleotide sequence analysis and advantages of automatic gene sequencers.
2. Localization of cloned segments in genomes – molecular and chromosomal location
3. Methods for determination of copy number of a cloned gene in genome
4. Mutant construction: Introduction, deletion, insertion and point mutation

### UNIT V

1. Principles and applications of Blotting techniques – Southern, Northern, Western and South-Western; Polymerase Chain reaction, Oligonucleotide synthesis, site directed mutagenesis, in-situ hybridization,
2. Principle and applications of Gel Mobility Shift Assay, DNA Fingerprinting and DNA Foot printing, Restriction fragment length polymorphism, Chromosome mapping and chromosome painting
3. Applications of Recombinant DNA Technology in Medicine
4. Biosafety measures and Regulations for genetically engineered products

### **Practical Exercises**

1. Bacterial Culture and antibiotic selection media. Preparation of competent cells
2. Isolation of plasmid DNA
3. Isolation of phage DNA
4. Quantitation of nucleic acids
5. Restriction mapping of plasmid DNA
6. Cloning in plasmid/phagemid vectors
7. Preparation of helper phage and its titration
8. Preparation of single stranded DNA template
9. Gene expression in *E. coli* and analysis of gene product
10. Polymerase Chain Reaction

### **Reference Books**

1. Recombinant DNA – By Watson et al
2. Principles of Gene Manipulation, Old and Primrose
3. Gene Cloning: An introduction , Brown
4. Biotechnology: Theory and Techniques (Vol I & II, 1995), Chirikjian
5. Molecular Genetics of Bacteria , Dale
6. Molecular Cloning (Vol I, II & III, 2001), Sambrook & Russell
7. Applied Molecular Genetics (1999), Miesfeld
8. Genes and Genome (1991), Singer & Berg
9. Molecular Biotechnology , Glick & Pasternak
10. Plant Molecular Biology (Vol I & II, 2002), Gilmartin & Bowler

*Note:* All text books are of latest editions:

## 302 : PLANT BIOCHEMISTRY

### UNIT I

1. Specialized plant organelles : Cell plate, Cell wall- Chemical and physical composition, biosynthesis, primary and secondary cell walls, Plasmodesmata, Plastids- Types and functions, Importance of vacuoles and microbodies, Meristematic cells and root quiescent zone
2. Absorption, adsorption and transport of water and ions in plants
3. Translocation of inorganic and organic substances
4. Structure and biogenesis of organelles involved in photosynthesis in plants

### UNIT II

1. Chloroplast membrane and molecular organization of thylakoids, proton gradient and electron transfer in chloroplasts of plants and in purple bacteria-difference from mitochondria
2. Light receptors- Chlorophyll, light harvesting complexes, bacteriorhodopsin as ion pump
3. Photosystem I and II- Location, mechanism of energy transfer between photosystems, ferredoxin, plastocyanin, plastoquinones and carotenoids
4. Hill reaction and photophosphorylation

### UNIT III

1. The Calvin Cycle- Evidence, mechanism and stoichiometry, role of light in activation of dark phase enzymes
2. Photorespiration : Mechanism and regulation
3. The C4 mode of photosynthesis : Mechanism, stoichiometry and purpose, difference from C3 in relation to plant productivity
4. Crassulacean Acid Metabolism : Mechanism and regulation

### UNIT IV

1. Biological Nitrogen Fixation : Formation of ammonia, conversion of nitrate to ammonia, assimilation and secondary assimilation of ammonia, inhibitors
2. Molecular properties of nitrogenase system, *nif* genes and their regulation, applications of biological nitrogen fixation
3. Molecular effects and mechanism of action of Auxin and Gibberellic Acid
4. Molecular effects and mechanism of action of Auxin and Gibberellic Acid

### UNIT V

1. Secondary metabolites : Plant alkaloids- Distribution, localization and biosynthesis of true and pseudoalkaloids, biological functions
2. Plant phenolics : nature, distribution, biosynthesis and regulation of phenolics, phenolic acids, hydroxycinnamic acid, phenylpropanes, quinones, xanthones, stilbenes, flavanoids, bioflavonoids, lignins and tannins, biological functions
3. Biochemistry of seed development and fruit ripening
4. Defense system in plants

### Practical Exercises

1. Estimation of plant proteins
2. Estimation of plant lipids and carbohydrates

3. Isolation of plant pigments, their analysis and determination of absorption Maxima
4. Chloroplast isolation
5. Hill Reaction
6. Estimation of nitrogenase
7. Estimation of nitrate reductase- *in vivo* method
8. Fruit ripening
9. Estimation of total phenolic compounds
10. Estimation of anthocyanin pigments

### **Reference Books**

1. Handbook of Photosynthesis (1997) by Mohammad Pe Sarakle
2. Plant Physiology by Salisbury and Ross
3. Introduction to Plant Biochemistry (1983) by Goodwin and Mercer
4. Seed : Physiology of Development and Germination (1994) by Bewley and Balck
5. Biochemistry of Energy Utilization in Plants (1987) by Blakie
6. Plant Biochemistry by Dey and Harbome

*Note:* All text books are of latest editions

## 303 : ANIMAL PHYSIOLOGY AND ENDOCRINOLOGY

### UNIT I

1. Composition of Blood : Structure, functions and biogenesis of RBC
2. Functions of hemoglobins, plasma proteins
3. Circulatory System (Open and closed circular, lymphatic systems)
4. Mechanism of blood coagulation : extrinsic and intrinsic pathways, inhibitors of coagulation

### UNIT II

1. Urine formation : Formation of dilute and concentrated urine
2. Regulation of water electrolyte balance and role of kidney and hormones
3. Acid-base balance and its regulation by kidney and hormones
4. Digestive system

### UNIT III

1. Nerve impulse transmission : structure of neuron, mechanism of conduction of nerve impulse along axon, neurotransmitters
2. Presynaptic and post-synaptic events of neuromuscular junctions
3. Ultra structure and molecular mechanisms of contraction of skeletal muscles and its regulation, energetics of muscle contraction, relaxation
4. Contraction of smooth muscles

### UNIT IV

1. General characters and classification of hormones, receptors and mechanism of action of hormones
2. Structure, synthesis, secretion, transport, metabolism and function of the hormones secreted by the pituitary
3. Structure, synthesis, secretion, transport, metabolism and function of the hormones secreted by parathyroid
4. Hormones of the thyroid structure, synthesis, secretion, transport, metabolism and functions

### UNIT V

1. Hormones of the adrenal medulla ; structure, synthesis, secretion, transport, metabolism and functions
2. Hormones of the adrenal cortex : structure, synthesis, secretion, transport, metabolism and functions
3. Hormones of the pancreas : structure, synthesis, secretion, transport, metabolism and functions
4. Hormones of the testis and ovary : structure, synthesis, secretion, transport, metabolism and functions

### Reference Books

1. Physiology by Guyton
2. Medical Physiology by Best and Taylor
3. Physiology by Garrett
4. Harper's Reviews of Biochemistry

*Note:* All text books are of latest editions.

## 304 : NUTRITIONAL AND CLINICAL BIOCHEMISTRY

### UNIT I

1. Basic Concepts : Composition of human body, Energy contents of foods, Measurement of energy expenditure, Direct and indirect calorimetry, Definition of BMR and SDA and factors affecting these, Thermogenic affects of foods, Energy requirements of man and woman and factors affecting energy requirements
2. Dietary requirements, sources of available and unavailable carbohydrates, physiological action of unavailable carbohydrates, Protein reserves of human body, Essential amino acids and concept of protein quality, Protein requirement at different stages of growth.
3. Nutritional and clinical significance of dietary calcium, phosphorus, magnesium, iron, iodine, zinc and copper.
4. Vitamins : Dietary sources, biochemical functions, specific deficiency diseases associated with fat and water soluble vitamins, Hypervitaminosis, Nutritional requirements during pregnancy, lactation and of infants and children

### UNIT II

1. Protein energy malnutrition : Etiology, clinical features, metabolic disorders and management of Marasmus and Kwashiorkar diseases
2. Obesity : Definition and classification, Genetic and environmental factors leading to obesity, Obesity related diseases and management of obesity, Role of leptin in regulation of body mass
3. Clinical nutrition : Role of diet and nutrition in the prevention and treatment of diseases, Dental caries, Fluorosis, renal failure, hyperlipidemia, atherosclerosis and rheumatic disorders
4. Food allergy : Definition, role of antigen, host and environment, Types of hypersensitivities, diagnosis and management of allergy

### UNIT III

1. Basic concept : Concept of accuracy, precision, normal and reference value determination, collection, processing and preservation of specimen, analysis, laboratory management, automation and quality control
2. Clinical significance of specific plasma/ CSF proteins, Clinical applications of serum protein eletrophoresis, Hemoglobinopathies
3. Carbohydrate metabolism : Carbohydrate intolerance, Diabetes mellitus, types, etiology and pathogenicity, Hypoglycemia, ketone bodies
4. Lipid metabolism : Diagnostic significance of analysis of serum lipids, chlolesteol and heart disease, Lipoprotein metabolism and disorders

### UNIT IV

1. Kidney ; Role of kidney in biochemical processes, Renal clearance, renal diseases and kidney function tests
2. Liver : Role of liver in biochemical processes, Bilirubin metabolism, ammonia metabolism, liver diseases and liver function test
3. Gastrointestinal tract : Disorders and diagnosis
4. Clinical enzymology Use of enzymes in diagnosis, Tissue distribution of enzymes, Diagnostic significance of acid phosphates, alkaline phosphatase, amylase, cholinesterase, creatine kinase, gamma glutamyl transfearse, lactate dehydrogenase, lipase

## UNIT V

1. Hemoglobin, porphyrin and related compounds : Disorders and diagnosis
2. Thyroid : Disorders and diagnosis
3. Hypothalamic-pituitary-adrenocortical system : Functions, disorders and diagnosis
4. Adrenal and gonads ; Disorder and biochemical assessment

### **Practical Exercises:**

1. Electrophoretic separation of serum proteins on agarose gel
2. Estimation of serum albumin and determination of albumin : globulin ration
3. Estimation of blood glucose by glucose-oxidase method
4. Estimation of serum triglycerides
5. Estimation of serum total cholesterol, HDL cholesterol, LDL cholesterol
6. Estimation of serum bilirubin
7. Electrophoretic separation of sickle hemoglobin
8. Estimation of serum acid phosphatase
9. Estimation of serum alkaline phosphatase
10. Estimation of serum aspartate transaminase
11. Estimation of serum creatinine

### **Reference Books**

1. Tietz Text book of Clinical Chemistry
2. Clinical Chemistry by DF Calbreath
3. Clinical Biochemistry by Varley

*Note:* All text books are of latest editions

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### **305 : Lab Course V**

Consists of practical exercises listed out under 301 & 302

### **306 : Lab Course VI**

Consists of practical exercises listed out under 303 & 304

## 401 : FRONTIER TECHNOLOGIES IN BIOSCIENCES

### UNIT I

1. Introduction and organization of animal cell and tissue culture laboratory, Contamination, Primary and established cell line culture
2. Serum and protein free defined media and their applications, measurement of viability and cytotoxicity
3. Introduction to balanced salt solutions and simple growth medium : Brief introduction of the chemical, physical and metabolic functions of different constituents of culture medium, Role of carbon dioxide and supplements
4. Biology and characterization of the cultured cells, measurement of parameters of growth

### UNIT II

1. Basic techniques of mammalian cell culture : Disaggregation of tissues, Primary and established cell line cultures and mass culture techniques
2. Stem cell technology : Types of stem cell, manipulations of stem cells
3. Factors governing manipulation of stem cell, Therapeutic cloning for embryonic stem cell (ESC's)
4. Applications of stem cell cultures

### UNIT III

1. Introduction of plant tissue culture and laboratory organization
2. Tissue culture media (composition and preparation), initiation and maintenance of callus and suspension culture, single cell clones
3. Organogenesis : Somatic embryogenesis, transfer and establishment of whole plants in soil
4. Protoplast isolation, culture, regeneration and somatic hybridization and applications of plant tissue culture

### UNIT IV

1. Nanobiotechnology : Introduction to nanoscience, Tools for measuring nano structures
2. Biosensor development and its applications
3. Microarray chips : Types of DNA chips and their production
4. SNP's and GMS (Genome mismatch repair)

### UNIT V

1. Functional proteomics : Methods of proteome analysis
2. Human Genome Project (HGP) : The human genome/ Social implications
3. Forensic applications of DNA analysis
4. Patents and intellectual property : Intellectual property areas : Trademarks, Copyrights. The process of obtaining a patent, Why obtain a patent, Recent changes in IPR and Patent policies

### Reference Books:

1. DNA Microarrays and Gene Expression by P.Baldi and G.W.Hatfield
2. Protein-Protein Interactions by Erica Golemis
3. A passion for DNA (Genes, Genomes and Society) by J.D.Watson
4. Modern Genetic Analysis by Anthony J.F. Griffiths et al.
5. Nanobiotechnology- Next Big Idea by Mark, Ratner, Daniel Ratner
6. Gene Cloning by T.A.Brown
7. Latest information on academic Websites

*Note:* All text books are of latest editions.

## **402 : BIOSTATISTICS, COMPUTERS AND BIOINFORMATICS**

### **UNIT I**

1. Introduction to Biostatistics, Common terms, notations and applications, Statistical population and sampling methods
2. Classification and Tabulation of data, Diagrammatic and graphical presentation
3. Frequency distribution and measures of central value
4. Measures of variability, Standard deviation, Standard error, Range, Mean, Deviation, Coefficient of variation

### **UNIT II**

1. Correlation and Regression, Positive and negative correlation, Calculation of correlation coefficient and regression coefficient, linear regression and regression equation
2. Test of significance; t-test, chi-square test and analysis of variance
3. Design of experiment, randomization, replication, local control, complementary randomized, randomized block design
4. Factor analysis, Path analysis

### **UNIT III**

1. Introduction to computer basics, Concept of Hardware and Software, DOS, Internal and External commands
2. Concept of file, folders, directories and their management
3. Office applications : MS-Office, MS-Word, MS-Excel and MS-Power Point
4. Open office on Linux : Word Processor, Spreadsheets, Impress

### **UNIT IV**

1. Overview of Bioinformatics : Introduction to MEDLINE on PubMed System for accessing Biological Information, Entrez, Swissport, PIR, NCBI
2. Sequence databases : Contents, Structure and annotation for Human Genome Databases, Plant Genome Databases, Retrieving and installing a program (Tree Tool), Multiple sequence alignment programme-Clustal W, X.
3. Use of genome analysis programs : BLAST, FASTA
4. CGC, Motif and Profile, Sequence search

### **UNIT V**

1. Predictive methods : Predictive methods using nucleotide sequences and protein sequencing, physical properties based on sequences, secondary structures, folding classes, tertiary structures
2. Genome mapping applications : EST and functional genomics, EST clustering, gene discovery, ORF prediction, molecular modeling
3. Phylogenetic analysis : Phylogenetic reconstruction, distance matrices, Parsimony, Phylips
4. Submitting DNA sequences to database

### **Reference Books:**

1. Introduction to Bioinformatics : A theoretical and practical approach by Stephen a Krawetz and DD Womble
2. Bioinformatics, Genes, Proteins and Computers by CA Orengo, DT Jopnes and JM Thornton
3. An Introduction to Computational Biochemistry by C.Stan and T.Sai

4. Instant Notes on Bioinformatics by DR Westhead, JM Parish and RM Twyman
5. Statistics for Agricultural Sciences by G Nageswara Rao
6. Fundamentals of Statistics by Goon et al.
7. Molecular Biotechnology : Therapeutic applications and Strategies by S Maulik and SD Patel
8. Methods in Biostatistics by BK Mahajan
9. Statistical Methods by SP Gupta
10. Statistical Methods by GW Snedecor and WG Cochran

*Note:* All text books are of latest editions.

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#### **403 : Lab Course VII**

Consists of practical exercises based on 401 & 402

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#### **404 : Research Project Work**

To be carried out on a specific defined objective under the supervision of a teacher. The work is compiled in the form of dissertation.

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